

COMPARATIVE HISTOLOGICAL AND HISTOCHEMICAL STUDY OF THE PAROTID GLAND IN BUFFALOES AND COWS

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ABSTRACT

The present work designed to describe the histological and histochemical features of parotid gland in buffaloes and cows. The glands were collected from ten heads of buffaloes and cows used. The histological and histochemical study revealed that the parotid gland in buffaloes and cows are compound tubuloacinar-type, having dense connective tissue capsule, the parenchyma consists of acini, intercalated, striated and excretory duct, acini surrounded by myoepithelial cells. The parotid is a purely serous gland. Histochemical study demonstrated mucopolysaccharide by using combined AB-PAS, acini shows weak to moderate reaction in buffaloes but weak reaction in cows.

INTRODUCTION

Salivary glands are extremely complex organs (1). They develop at different locations, they have different architectures and produce more than one type of saliva (2). Saliva in the ruminant is primarily supplied by parotid glands, which secrete rapidly and continuously (3). This is highly important for the animals, because they need to relocate nitrogenous and phosphorous compounds, which are essential factors for microbial growth in the forestomach (4). In contrast to humans, the ruminant secretes large volumes of alkaline and well buffered saliva, mostly for lubricating and swallowing food particles (5). Salivary glands are important for research because of their diverse functions (6).

The present study aims to describe histological and histochemical feature of parotid gland in buffaloes and cows.

MATERIALS AND METHODS

Twenty healthy adult heads(10 buffaloes and 10 cows) were bought from the slaughter house at Basra province.The parotid glands were removed directly then serial processes were done according to (7).Later the sections were stained with Harris haematoxylin and eosin, Masson's trichrome stain for appearance of connective tissue, Van Gieson stain differential of collagen and other connective tissue and in the histochemical study AB(PH2.5)-PAS method for mucosubstances. The Statistical analysis was performed on the basis of T-Test by using a Statistical program as described by (SPSS,2012,version21).

RESULT

The microscopic structure of parotid gland in buffaloes and cows revealed that the gland is surrounded by dense connective tissue capsule, septa extend from the capsule and separated the gland into many lobules with different shapes and size, these septa are characterized by the presence of adipose tissue (Fig 1,2).The lobules are composed of pure serous acini which were small in diameter and had small lumen which was difficult to distinguish.The statistical analysis revealed that the mean diameter and lumen of acini were $(36.50 \pm 0.45, 8.50 \pm 0.61) \mu\text{m}$, respectively in buffalo and $(35 \pm 0.95, 5.5 \pm 0.5) \mu\text{m}$ respectively in cow (Table1).The acini was characterized by pyramidal shaped lining cells with round centrally located nuclei, very fine and scanty connective tissue were found between acini as intralobular septa, the myoepithelia cells were seen surrounding the acini(Fig.3,4).Intralobular duct system which was initiated by intercalated duct and are lined by simple cuboidal epithelium and elongated nuclei (Fig.3,4).These duct was emptied into larger striated duct, the latter duct are lined by simple columnar epithelia and characterized by acidophilic cytoplasm and large centrally located nuclei.These duct are surrounded by fine connective (Fig.5,6).The intralobular striated duct are converged together to form the larger interlobular ducts, which are lined by simple columnar epithelium cells at the beginning and change to stratified epithelia when they are increase in size.These duct located in the interlobular septa (Fig. 7,8).The large interlobular ducts are connected together to form the main excretory ducts of gland which are lined by stratified

columnar epithelia and goblet cells. These ducts were enclosed by connective tissue stroma (Fig.9,10). Histochemically the section from parotid gland stained with (AB(PH2.5)-PAS) revealed weak to moderate reaction in serous acini in buffalo and weak reaction in serous acini in cow and all type of duct show negative reaction only goblet cell were positive toward stain.

Table (1): The Mean for diameter and lumen of secretary end pieces, Intercalated, striated and interlobular duct for parotid salivary gland in buffaloes and cows

| Gland Name | Side | Mean ± S.E | Mean ± S.E | Mean ± S.E | Mean ± S.E | Mean ± S.E | Mean ± S.E | Mean ± S.E | Mean ± S.E | T-Test |
|--------------------|--------------|--------------|------------|-------------|-------------|------------|--------------|------------|-------------|--------|
| | | Dia. (µm) | Lumen (µm) | Dia. (µm) | lumen (µm) | Dia. (µm) | Lumen (µm) | Dia. (µm) | Lumen (µm) | |
| Parotid in Buffalo | Serous Acini | 36.50 ± 0.45 | 8.5 ± 0.61 | 52 ± 1.46 | 32.5 ± 1.77 | 70 ± 0.71 | 52.50 ± 0.94 | 185 ± 0.79 | 97.5 ± 0.12 | N.S |
| Parotid in Cow | Serous Acini | 35 ± 0.95 | 5.5 ± 0.50 | 62.5 ± 0.59 | 32 ± 0.42 | 80 ± 0.90 | 50.50 ± 0.82 | 150 ± 1.07 | 98.9 ± 0.58 | N.S |

SE: Standard Error Mean, N.S: Mean there is no difference between groups at (P>0.05)

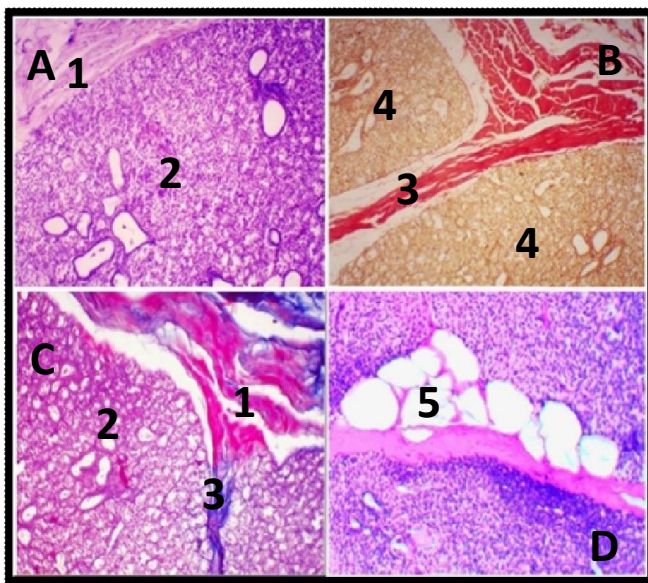


Fig. (1) Section from parotid salivary gland in buffalo showing: A-1. capsule, 2.acini (H&E 100x), B-3.septa, 4.lobules (Van Gieson 100 X), C-(masson's trichrome stain100x), D-5.adipose tissue (H&E 100x).

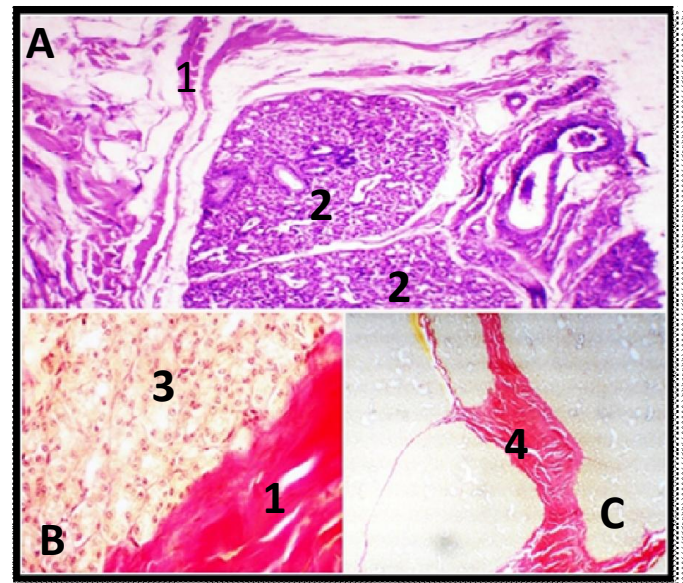


Fig. (2) Section of parotid salivary gland in cow showing: A-1.capsule, 2.lobule (H&E40x), B-3.acini (Van Gieson 400x), C-4.septa (H&E 100x).

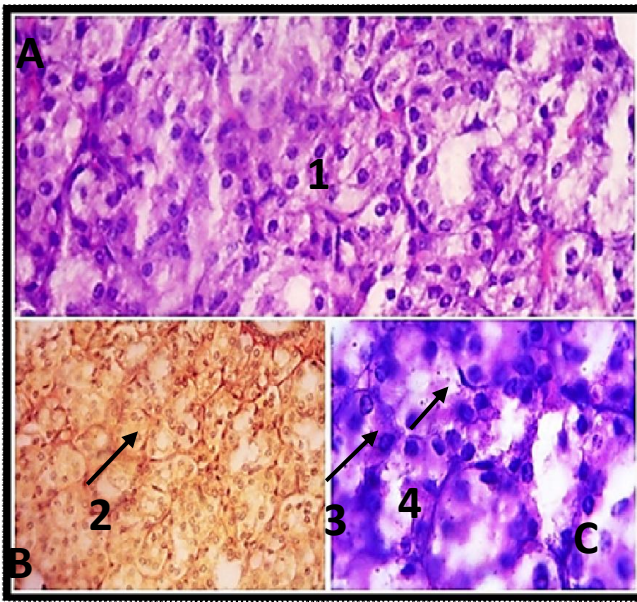


Fig. (3) Section from parotid salivary gland in buffalo showing: A-1.Serous acini (H&E 400x), B-2.lumen of acini (Van Gieson400X), C-3.Pyramidal cell of acini, 4.Myoepithelia cell (H&E oil immersion).

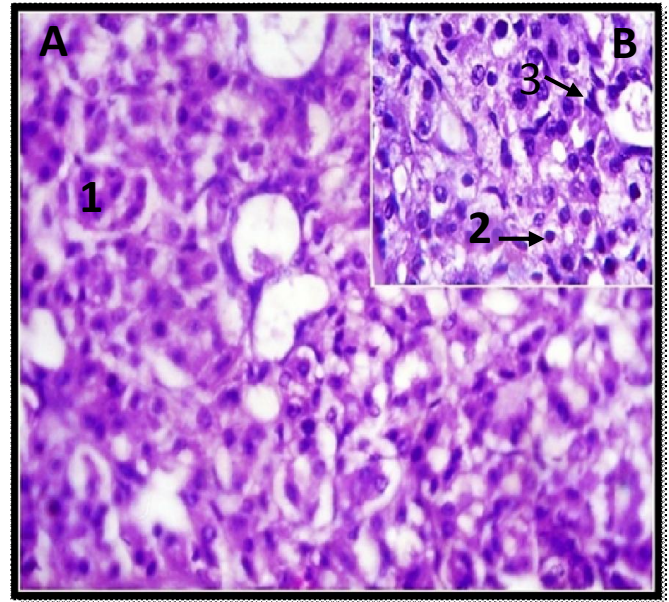


Fig. (4) Section of parotid salivary gland in cow showing: A-1.Serous acini (H&E400x), B-2.Cuboidal cell of acini, 3.Myoepithelia cell (H&E oil immersion).

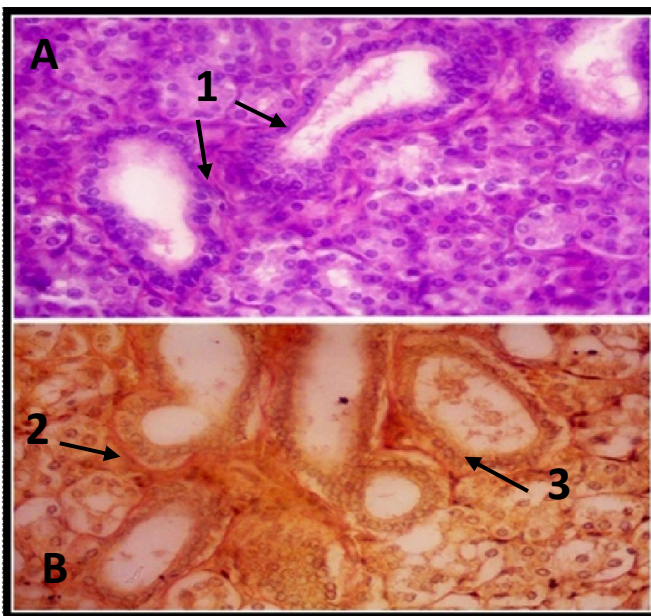


Fig. (5) Section of parotid salivary gland in buffalo showing: A-1.Striated duct (H&E 400X), B-2.Simple columnar epithelia of striated duct, 3.Intercalated duct (Van Gieson stain 400x).

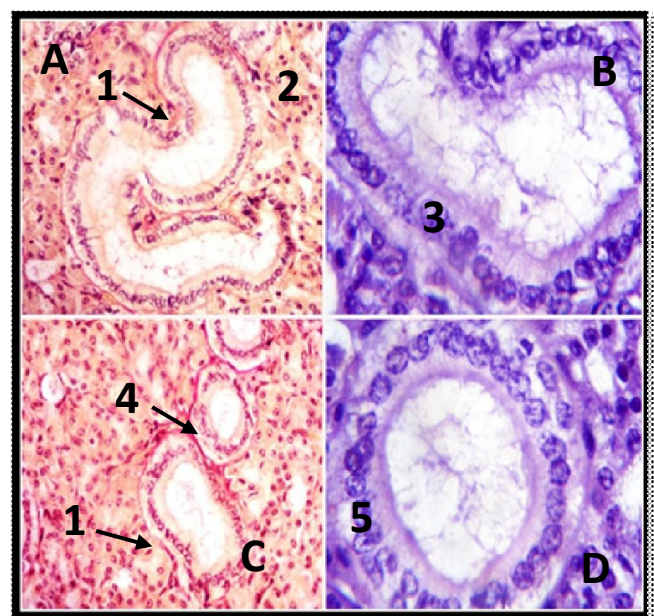


Fig. (6) Section of parotid salivary in cow showing: A-1.Convoluted appearance of striated duct, 2.Acini (Van Gieson stain 400x), B-3. striated duct with simple epithelia (H&E oil immersion), C-4.intercalated duct (Van Gieson stain 400x). D-5. intercalated duct with simple cuboidal (H&E oil immersion).

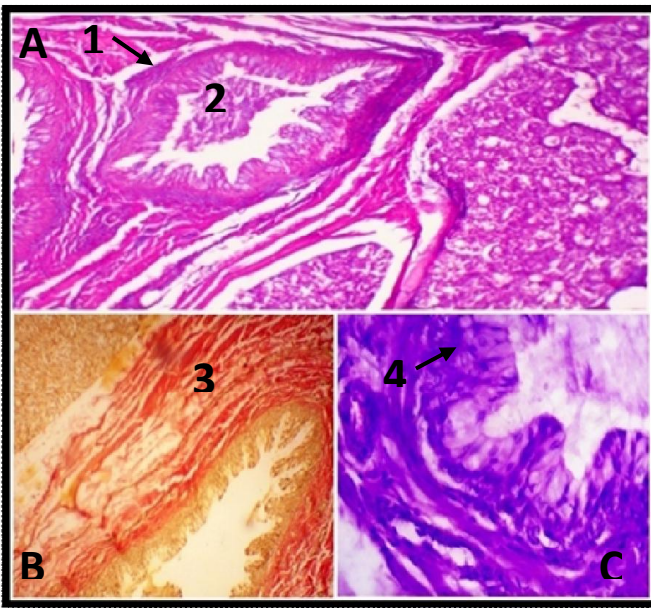


Fig. (7) Section of parotid Salivary gland in buffalo showing: A-1.Interlobular duct, 2.lumen with secretion (masson's trichrom stain 100x), B-3.Interlobular connective tissue around the duct (Van Gieson 100x), C-4.Stratified epithelia of interlobular duct (H&E 400x).

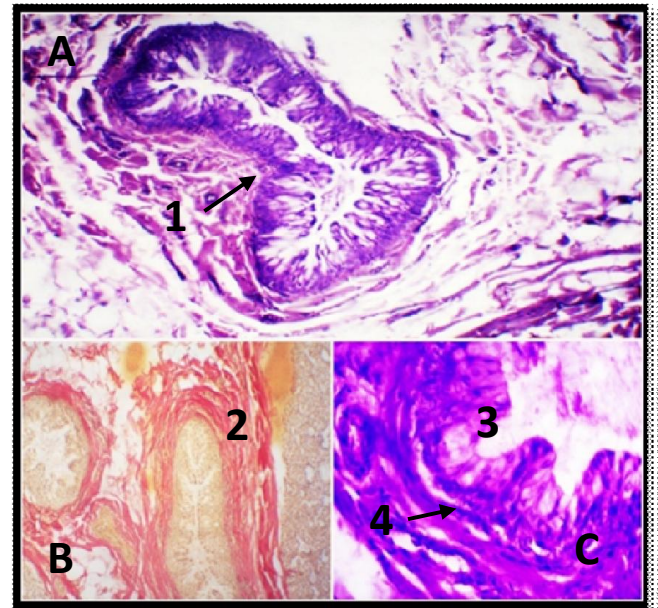


Fig. (8) Section of parotid salivary gland in cow showing: A-1.Interlobular duct (H&E 100x), B-2.Fibrous tissue around the duct (Van Gieson stain 100x), C-3.stratified epithelia, 4.Goblet cell (H&E 400x).

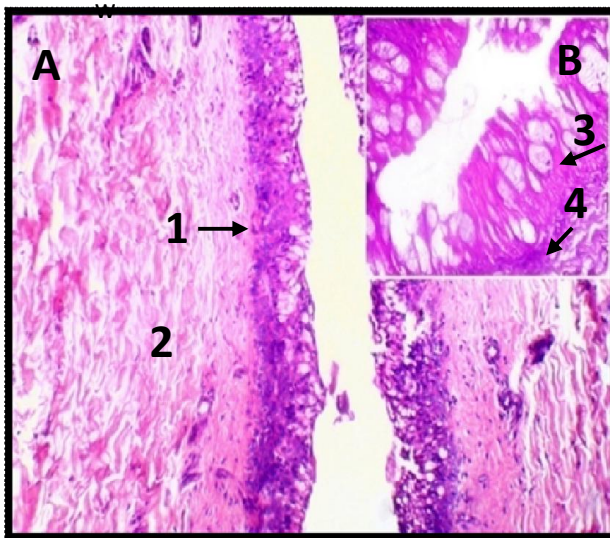


Fig. (9) Section in parotid salivary gland of buffalo showing: A-1.Main excretory duct, 2.fibrous tissue (H&E100X), B-3.Vacuolated epithelia, 4.Goblet cell (H&E 400x).

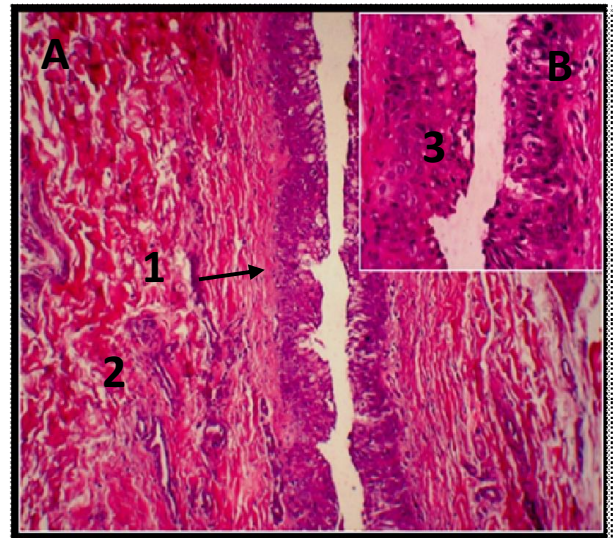


Fig. (10) Section of parotid salivary gland in cow showing: A-1.Main excretory duct, 2.Fibrous connective tissue (H&E 100x), B-3.Stratified epithelia of duct (H&E 400x).

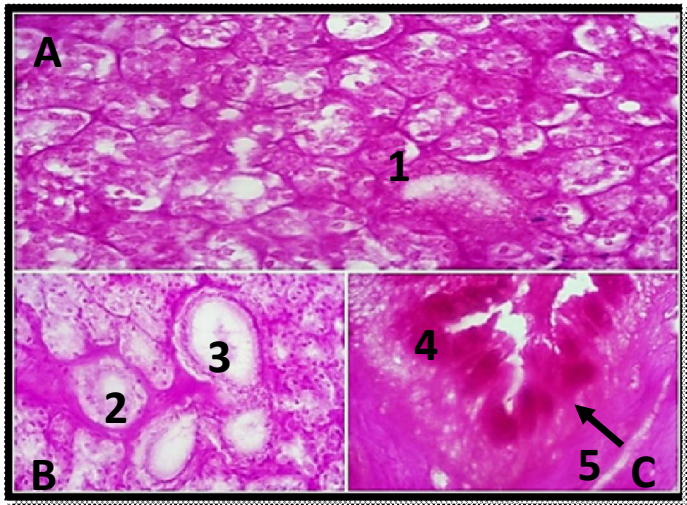


Fig. (11): Section of parotid salivary gland in buffalo showing: A-1.Serous acini (AB-PAS 400x), B-2.Intercalated duct, 3.Striated duct (AB-PAS 400x), C-4.Interlobular duct, 5.Goblet cells (AB-PAS 400x).

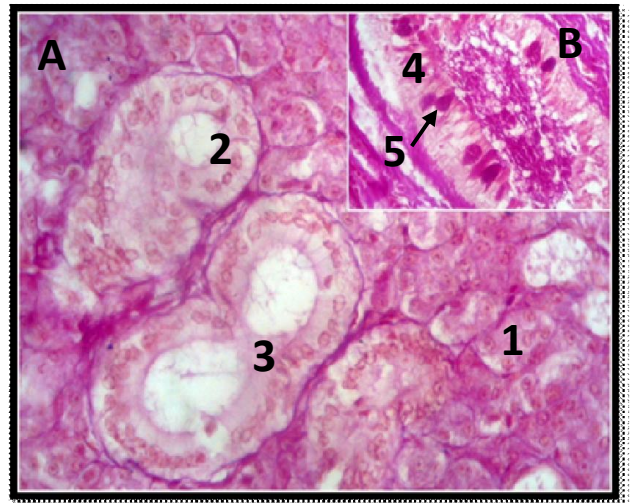


Fig. (12) Section of parotid salivary gland in cow showing: A-1.Serous acini, 2.Intercalated Duct, 3.Striated duct (AB-PAS 400x), B-4.stratified epithelia of Interlobular duct, 5.Goblet cells (AB-PAS 400x).

DISCUSSION

The parotid gland in cattle and buffalo were compound tubulo-alveolar gland this finding was agreement with (8,9,10,11).The gland was covered by capsule of dense regular connective tissue this result was agreement with (12). (13), who reported that covering capsule is used for protecting the serous secretory acini cells, the observation of serous acini had been observed by (14),(8), (15), (16),(17), (18), (19), (20), (21), (22). The finding was disagreed with(23), (24), (25),(26),(27), they found that the parotid gland was mixed gland serous and mucous.(28)mentioned that the serous type was found in the parotid gland of herbivorous animal and mixed in the carnivorous related to the type of feeding of these animal.

The occurrence of well-developed serous cells may be an adaptation for raise digestion of carbohydrates in the oral cavity and also to increase production of antibacterial agent lysozyme to help reduce rate of infection establishment in the wild (29).The present study showed that myoepithelial cells surrounding the serous acini this finding was agreement with (30) while different with (31) the serous acini lack

myoepithelial cells and only cytoplasmic processes of cytoplasmic processes of myoepithelial of intercalated duct were found between acini.

The myoepithelial cells surrounding the acini secretory end piece was provides contractile force to help move secretion from the acinicells and push them during the intercalated duct (32).The intercalated duct of simple cuboidal epithelium play important role in transport secretion from acini to striated duct, this simple cuboidal was reported by (22).The striated duct was lined by simple columnar epithelium it was transport secretion from the intercalated duct to the excretory duct,A tall cuboidal epithelium of striated duct was reported by(33), this result disagree with (34) reported that the intralobular duct in buffalo without striated duct.The interlobular duct was transport product to the oral cavity which was lined by stratified epithelium may reveal the need for protection of underlying basement membrane by occasional action of activated serous fluid enzyme (13).The distribution of goblet cells with epithelium of main duct play important role in modulation of saliva secret from acini by addition of mucous from goblet cells (8). The acini small in diameter and lumen this agreement with (8)In the histochemical study the parotid gland of buffalo show weak to moderate reaction with combined AB-PAS(PH 2.5)and in cow the acini show weak reaction,this result was confirmed by (8),(34) who mentioned that the secretions of the parotid gland contained neutral mucins and some sialo mucins.

Conclusion:

The parotid gland in both buffalo and cow have same structure and no significant difference between them, histochemically parotid gland in buffalo was seromucous gland.

دراسة نسيجية وكيمونسيجية مقارنة للغدة النكفية في الأبقار والجاموس

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الخلاصة

هدفت الدراسة الحاليه لمعرفة التركيب النسيجي والكيمونسيجي للغده النكفيه في الابقار والجاموس حيث تم جمع العينات من عشرون راسا" من الجاموس ومثلها من الابقار وجميع العينات بالغة وسالمة مظهريا. أظهرت الدرسة النسيجه ان الغدة النكفية مركبة نسيجية عينية تحاط بمحفظة من النسيج الرابط تكون عنيباتها

مصلية الافراز وقد ظهر الجهاز القنوي متكاملًا" والذي يتألف من القنوات البينية، والمخططة وبين الفصيصات. اما في الدراسة الكيمونسية فقد بينت الغدة النكفية تفاعلاً " ضعيفاً الى متوسط الشدة في العنبيات المصلية في الجاموس وتفاعل ضعيف في العنبيات المصلية في الابقار.

REFERENCE

- 1- Young, J. A. and Van Lennep, E.W. (1978). The morphology of salivary glands. Academic Press, pp:273.
- 2- Jaskoll, t.; Zhou, Y. M.; Chai, Y.; Makarenkova, H.P.; Collinson, J.M.; West, J.D. and Carvalho, A.D. (2002). Embryonic submandibular gland morphogenesis: stage-specific protein localization of FGFs, BMPs, Pax6 and Pax 9 in normal mice and abnormal phenotypes in FgfR2-IIIc(t/Delta), BMP(-/-) and Pax(-/-) mice cells tissue organs. 170: 83-90.
- 3- Piatkowski, B.; Gurtler, H. and Voigt, J. (1990). Grundzüge der Wiederkäuer – Ernährung 1. Auflage, Gustav Fischer – verlag, Jena Germany. PP:59-67
- 4- Breves, G. ; Rosenhagen, C. and Hoeller, H. (1987). Saliva secretion of inorganic phosphorus in phosphorus-depleted sheep. J.V.Med; A43,42-47.
- 5- Kay, R.N.B. (1960) The rate of flow and composition of various secretions in sheep and calves. J. Physiol. 150, 515-537.
- 6- Asari, M.; Kimura, H.; Chihara, N.; Kasuya, T. and Nishita, T. (2000). Immunohistochemistry of carbonic anhydrase isoenzyme (CA-I, II and III) in canine salivary glands. A distributional and comparative assessment. J. Vet. Med., 29:9-12.
- 7- Bancroft, J.D and Stevens, A. (2012). Theory and practice of histological techniques, 7th edition. Churchill Livingstone. pp: 127-129.
- 8- Al-Saadi, A.I. (2002). Anatomical, histological and ultrastructural study of the major salivary glands in the endogenous buffalo. (M.Sc. Thesis), college of veterinary medicine, university of Baghdad.
- 9- Barnwal, A.K. and Sinha, R.D. (1985). Histological and histochemical studies on the secretory end-pieces of the parotid salivary gland of buffalo. Indian J. Anim. Sci., 55(1):1-4.

- 10- Al-Samarrae, N.S.; Rabie, F.O. and Abass, T.A. (1989). Topography and histology of parotid gland in one-humped camel. *Iraq.J.Vet.*, 13:42-51.
- 11- Boshell, J.L.; Walter, H. and Wilborn (1978). Histology and ultrastructure of the pig parotid gland. *American. J. of ana*;152 (4):447-465.
- 12- Khojasteh S. M.B. and Delashoub M. (2012). Microscopic anatomy of the parotid and submandibular salivary gland in European hamster (*Cricetus cricetus* L.). *I.research j. of applied and basic science*. vol,3(7),1544-1548.
- 13- Ekele, I., Uchenna, N., Okechukwu, N. and Isaiah, A. (2013). Histology of the parotid salivary gland of the African Palm quirel. *Rev. Fac. Cs. Vets. University of Agriculture Umudike, Abia state ,Nigeria* ,54 (1) :11-16
- 14- EL-Ramli, A.; Yasear, A.Y.; Sultan, A. (2013). Structural Histological changes in the parotid salivary gland of rabbit treated with neostigmine. *J. Basic Med. Allied Sci.*, 1:1-15.
- 15- AL-Saffar, F.j. and Simawy, M.S.H. (2014). Histomorphological and histochemical study of the major salivary glands of adult local rabbits. *Int. J.Adv, research*. V(2), (11). 378-402.
- 16- Kim ,S.K. (1976). The cytochemical localization of adenylate cyclase activity in mucous and serous cells of the salivary gland .*J.Supramolecular structure* 4:185(145)-197(157).
- 17- Stephens, L. C., King, G. K., Peters, L. J., Ang, K. K., Schultheiss, T. E. and Jardine, J. H. (1986). Unique radiosensitivity of serous cells in Rhesus monkey submandibular glands. *A. J. P.*, 124 (3): 479-487
- 18- Al-Okaili, A. G., Sedeeq, B. I., Hazeem, M. I. (2008). Histological changes of the submandibular Salivary gland of mice maintained on a liquid diet. *College of Dentistry, Tikrit University*.,1- 4
- 19- Elewa, Y.H., Bareedy, M.H., Abuel-Atta, A.A., Ichii, O., Otsuka, S., Kanazawa, T., Lee, S., Hashimoto, Y. and Yasuhiro Kon, Y. (2010). Structural characteristics of goat (*Capra hircus*) parotid salivary glands. *Jpn. J. Vet. Res.*, 58(2): 121-135
- 20- zhou, J., Wang, H., Yang, G., Wang, X., Sun, Y., Song, T., Zhang, C. and Wang, S. (2010). Histological and Ultrastructural characterization of developing miniature pig salivary glands the *Anat.record.*, 293:1227-1239.
- 21- Adnyane, I.K.M., Zuki, A.B., Noordin, M.M. and Agungpriyono, S. (2010). Histological study of the parotid and mandibular glands of barking deer

- (*Muntiacus muntjak*) with special reference to the distribution of carbohydrate content. *Anat. Histol. Embryol.*, 39: 516–520
- 22- Amano, O.; Mizobe, K.; Bando, Y. and Sakiyama, K. (2012). Anatomy and histology of rodent and human major salivary glands. *Acta Histochem. Cytochem*; 45 (5): 241- 250.
- 23- Singh ,S. ;Pawar,HS. ; and Roy ,K .(1995) .Histoenzymic study on parotid salivary gland of castrated donkey .*Indian .J.Sci .* ,65(2) :137-139.
- 24- Condon, S.M.; Estecondo,S. and Casanave, E.B. (2003). Histological study of the salivary glands in *Dasyprocta hybridus*(Mammalia,Dasyproctidae) *Int.J. Morphol.*, 21(3):199-204.
- 25- Van Lennep, E. W. and Young, J. A. (1978). *The Morphology of Salivary Glands*mic Press; London PP: 135-140
- 26- Leone,A.; Spatola,G .F.; Cucco, D.; Tessitore, V.;Bonaventura, G.andUzzo, M.L. (2012). Immunohistochemical expression and distribution of orexin,orphanin and leptin in the major salivary glands of some mammals. *Folia.histo. et cytobiologica*.50(4) pp;504-512.
- 27- Poddar,S. and Jacob,S.(1977) Gross and microscopic anatomy of the major salivary glands of the ferret .*Acta .Basel* ,98(40) :439-443.
- 28- Elewa, Y. H. A., Ichii, O., Otsuka, S., Hashimoto, Y. and Kon, Y. (2014). Structural change of goat parotid salivary gland: pre- and -post weaning periods. *Anat. Histol. Embryol.*, 43(4): 265-272.
- 29- Ognean, L.; Dojana, N.; Corina, R.; Fiziologia, A. (2000). Ed. *Presă Universitară Clujeană*, (II), ClujNapoca.
- 30- Suzuki ,S. ;Ago ,A.; Mohri,S. *et al* .,(1983). Fine structure of parotid gland of Djungarian hamster (*Phodopus sungarus*) *Jikken Dobutsu* ,23(4) : 175-184.
- 31- Redman, R.S., Sweney, L.R., McLaughlin, S.T. Differentiation of myoepithelial cells in the developing rat parotid gland. *Am J Anat.* 1980;158:299–320.
- 32- Redman, R. S. (1994). Myoepithelium of salivary glands. *Microsc. Res. Techn.*, 27: 25- 45.
- 33- Bazan, E.; Watanabe, I.; Iyomasa, M.M.; Mizusaki, C.I.; Sala, M.; Lopes, R.A. 2001. Morphology of the submandibular gland of the gerbil (*Meriones unguiculatus*). A macroscopic and light microscopy study. *Chil. Anat.*,PP: 19.

- 34- Pal ,C. , Candra,G. and Bharadwaj ,M.B. (1971) .Histological and certain histochemical observation on the parotid salivary gland of buffalo *Bubalus bubalis* .Indian J .Anim.Sci.,42:420 426.