See discussions, stats, and author profiles for this publication at: https://www.researchgate.net/publication/310624761

Synthesis, Spectroscopic Characterization, Thermal Stability and antimicrobial activity of Schiff base, β-lactam and Zn (II), Cu (II) complexes derived from Sulfamerazine.

reads 30

a
Ali Ghanim
University of Basrah
3 PUBLICATIONS 7 CITATIONS
SEE PROFILE

Some of the authors of this publication are also working on these related projects:

Project inclusion complexes View project



Research Journal of Pharmaceutical, Biological and Chemical Sciences

Synthesis, Spectroscopic Characterization, Thermal Stability and antimicrobial activity of Schiff base, β- lactam and Zn (II), Cu (II) complexes derived from Sulfamerazine.

Jabbar S Hadi^a*, Abdulelah A Almayah^b, and Ali G Swadi^a

^aCollege Of Education For Pure Science-Basrah University-Basrah-IRAQ ^bCollege of Pharmacy-Basrah University-Basrah-IRAQ

ABSTRACT

Schiff base derived from *o*-vanillin and Sulfamerazine, β – lactam and Zn(II), Cu(I) complexes have been Synthesized and characterized by IR, ¹HNMR, MASS spectrometry, molar condactance and thermal analysis. The schiff base acts as a monobasic bidentate ligand in complex formation thermal analysis indicate the presence of lattice water molecules in complexes. The molar condactane measurments indicate the non electrolyte behaviour of the complexes in DMF solution. The antimicrobial activities of compounds were tested against four bacterial clinical isolates (human pathogenic) strains as 1 gram +Ve bacteria (*Staphylococcus aureus*), *3 gram* – *Ve bacteria(Escherichia coli , Pseudomonas aeruginosa* and *Proteus spp.*) to develop novel class of anti microbial agents with varied mode of action. The results of bioassay showed that the newly synthesized β -lactam emerged as laed with MIC (mg/ml) values with mentioned gram +Ve,While the complexes-schiff base showed highly antimicrobial activity toward mentionded gram –Ve bacterai. These results compared with standard drugs (Cephalexin;30µg/disc, Ciprofloxacin;5µg/disc, Oxacillin;1µg/disc, Cloxacillin;1µg/disc, Aztreonam;30µg/disc, Ampicillin;10µg/disc, Clarithromycin;15µg/disc , Novobiocin;30µg/disc)

Keywords: Sulfamerazine , antimicrobial activity , β – lactam , Thermal stability.

*Corresponding author



INTRODUCTION

Schiff bases have recieved much attention due to their Characteristic properties and applications such as structural variety [1], Varied coordinating ability [2,3], thermal Stability [4,5], Catalysis properties [6,7]. biological activities [8-10]etc . Schiff base derived from sulfa drugs have gained considerable importance due to their pronounced biological activities .Schiff base derived from sulfa drugs are good ligands due to the presence of different active site such as heterocyclic moieties NH, OH and SO2 groups, this led to the formation a stable complexes with d-block transition metals with a wide range of applications. β – lactam are synthetic antibiotics active against Gram-positive and Gram-negative bacteria by inhibiting the synthesis of the peptidoglycan layer from the cell wall [11]. The highly stained β – lactam nucleus is stabilized by means of the fusion of a variety of either 5-membered or 6-membered hetetocyclic moieties to give rise to a wide spectrum of newer antibiotics [12].

EXPERIMENTAL

Materials

Analar grade Cu $(CH_3COO)_2$. H_2O and $Zn(CH_3COO)_2$. $2H_2O$ as well as o.vanillin were obtained from fluka. Sulfamerazine and chloroacetylchloride were obtained from sigma and used as recieved. Muller-Hinton agar, Nutrient agar, Mckonky agar and Nutrient Brouth were obtained from HiMEDIA. Standard drugs were obtained from Bioanalyse.

Instrumentation and spectral measurments

Meliting points were recorded by using a Fisher Johus melting pointapparatuse IR spectra were recorded by using shimadzu FTIR – 8300 spectrophotometer in the region 4000-400cm⁻¹ in KBr pellet. The Mass spectra were scanned by EI technique at 70eV with an Agilent technologies 5975C spectrometer. ¹HNMR spectra were scanned on a Bruker 400MHz spectrometer TMS as the internal standard and DIMSO-d₆ was used as solvent. The molar condactance of complexes was measured using conductometer coming model 441at room temperature using DMF as a solvent. The thermal analysis (TG and DTG) were carried out in dynamic Argone atmospher (20ml/min) witha heating rate of 10°C/min using a Rheometric scintific and TGAQ50(USA) .Sterlizer Aesthtica UV.Ray (USA) .Incubator (HI 900 D) at 37°C. Autoclave (HIRAYAMA) at 120°C.

Synthesis

Synthesis of schiff base ligand (A₁):4-(2-hydroxy-3-methoxybenzylideneamino)-N-(4-methylpyrimidin-2-yl) benzenesulfonamide.

2 mmole(0.538g) of sulfamerazine (S.M) dissolved in 10ml ethyl actate was mixed with 2mmole (0.304) of o.vanillin dissolved in 10ml absolute ethanol.2 drops of H_2SO_4 was



added as a catalyst. The resulting mixture was reflexed for 2hrs. The orange solid preciptate which formed during the raection filtered hot and washed with cold ethanol. The orange crystal product is produced in 75% yield and melting at 202-203°C.

Synthesis of β -lactam (A₂) :4-(3-chloro-2-(2-hydroxy-3-methoxyphenyl)-4-oxoazetidin-1-yl)-N-(4-methylpyrimidin-2-yl)benzenesulfonamide.

1mmole of (A₁) dissolved in 10ml of Dioxane and the solution was kept at 0-5°C in ice water. Then, 1mmole (0.14ml) of triethylamine was added and the mixture stirred for 5 min. Then, 2mmole (0.16ml) of chloroacetyle chloride was added dropwise and finally the mixture was stirring for 4hrs and left at room temprature over night. The mixture was poured into crush ice and the white solid preciptate was filtered and washed with ethanol,dried at 70°C .yield 37% and m.p 190°d.



(A₂) B-Lactam

Scheme 1: preparation of (A₁)and(A₂)



Synthesis of Zn complexes (ZnA₁) and Cu complexes (CuA₁):

Metal complexes ZnA₁ and CuA₁ were prepared using template reaction as follow. In methanolic solutions of each sulfamerazine (2mmole), o.vanillin (2mmole).The pH of the mixture was adjusted to 7.8-8 bu adding drops from 10%NaOH. Methanolic solution of metal acetate (1mmole in 10ml) then added to the reaction mixture drop wise with constant sterring and finally heated under reflex for 3hrs.The mixture was concentrated by evaporation. The precipitated metal complexes were filtered off, then washed with ether and preserved in desicator over anhydrous Calcium Chloride



Scheme 2: preparation of ZnA₁ and CuA₁

Bacterial Isolates

Four bacterial clinical isolates were obtained from laboratory research/Clinical laboratory science branch/Collage of Pharmacy/Basrah University, and they are: (*Staphylococcus aureu*, *Escherichia coli*, *Pseudomonas aeruginosa* and *Proteus spp.*)

July - August



Antimicrobial activity investigation

Disc diffusion by Baueyer-Kirby method [13] were applied .Circular disc of watman No.1 were sterilized by Ultra-Violet Ray (U.V). The discs papers were imprignated into prepared chemical compounds (S.M , A_1 , A_2 , Zn A_1 and Cu A_1) in one (1ml) DMSO as solvent with differant concetration (1mg ,10mg ,50mg ,100mg) .The clinical isolates were cultured and spreading on Mullar-Hinton agar,then discs were applied in each bacterial isolates .

Standard antimicrobial investigation

Standard antibiotic discs (Cephalexin, Ciprofloxacin, Oxacillin, Cloxacillin, Aztreonam, Ampicillin, Clarithromycin and Novobiocin) were used to comparison with prepared chemical compounds.

Statistical Analysis

Analysis of variance(ANOVA) [14] were used to explain the significancy differences of antimicribial activity among bacterial isolates.

RESULTS AND DISCUSSION

Schiff base (A₁) was prepared from condensation of Sulphamerazine(S.M) with o.vanillin and then coverted to β -lactam (A₂) by cycloaddition of chloroacetyle chloride with (A₁) (scheme 1). Two complexes(Zn complex) and (Cu complex) were prepared from A₁. The observed physical properties of compounds were collected in (Table 1).

compound	Chemical formula MW.g/mole	m.p °c or °d	physical state	molar conductance	Yield %
A ₁	C ₁₉ H ₁₈ N ₄ O ₄ S 398	202-203.5	orange crystal		75
A ₂	C ₂₁ H ₁₉ N ₄ O ₅ SCI 474	190	white powder		37
ZnA ₁	C ₃₈ H ₃₈ N ₈ O ₁₀ S ₂ Zn 896	>300	yellow powder	11	67
CuA ₁	C ₃₈ H ₃₆ N ₈ O ₉ S ₂ Cu 876	>300	brown powder	3	63

Table 1 : Physical properties of prepared compounds

Compound A₁ orange in colour, A₂ white colour ,air stable ,having sharp melting points >200°C . (A₁) soluble in dioxane,DMF and DMSO but, insoluble in common organic solvents. (A₂) soluble in acetonitrile,DMFandDMSO but, insoluble in most common organic solvents. Complexes (ZnA₁) and (CuA₂) are stable and decompose >300°C,they were insoluble in most common organic solvents but soluble in DMSO and DMF.



IR Spectra analysis

The IR spectrum of the free legand A_1 (Fig 1) shows broad band at 3462cm^{-1} that attributed to the hydrogen bonded (OH) group . A strong band is observed at 1616cm^{-1} which is assigned to ($C=N_{azo}$), this band is shifted to a lower wave number side $\Delta V = 4 \text{cm}^{-1}$ in case Zn(II) complexe and $\Delta V = 17 \text{cm}^{-1}$ in case of Cu(II) complexe (Fig2), which indicates the participation of the azomethine groups in coordination to the metal ions through the lone pair of electrons on the nitrogen[2,4]. In addition the involvement of phenolic OH group in bonding with metal ions can be examined by the (C-O) band which is shifted to lower wave number side $\Delta V = 23 \text{cm}^{-1}$ in case Zn(II) complexe and $\Delta V = 48 \text{cm}^{-1}$ in case of Cu(II) complexe. The IR spectrum of A_2 (Fig 3) shows a new band at 1687cm^{-1} that corresponding to ($C=O_{\beta-Lactam}$) [15,16] of lactam moity, this gives information regarding lactam ring formation as well the disappearance of $C=N_{azo}$.



¹HNMR Spectra Analysis

The ¹HNMR spectral data of the schiff base ligand (A₁) confirms the proposed structural elucidation of the ligand, this ¹HNMR spectrum(Fig 4)shows the methyl protons as a singlet signal at σ 2.4ppm, it also displays a singlet signal at σ 3.9ppm which is attributed to methoxy protons. The azomethine proton appear at σ 8.6ppm.The signal at σ 10.7ppm is attributed to –



NH proton [4] and the signal of –OH proton appear at σ 13.07ppm. A comparison of the chemical shifts of ligand A₁ with its Zn complex (Fig 5) indicates that the signal due to phenolic proton is absent in the complex spectrum ,this can be attributed due to the deprotonation of the phenolic group and subsequently the replacement of the proton by metal [2 ,4]. The azomethine signal is observed up field at σ 8.4 in complex spectrum, it supports the coordination of azomethine nitrogen to matal ion. β -lactam (A₂) formation is characterized by the disappearance of the azomethine proton signal of the corresponding schiff base (A₁) and appearance of the two new signal of the protons of β -lactam ring (Fig 6) .The chemical shift of proton HC-N appear at σ 3.57ppm [17] andHC-CI proton appear at σ 2.57ppm [18].The -OH proton signal which would have undergone very rapid exchange with the solvent appear as quit broad signal ato 4ppm [19 ,20] .











Figure 6: ¹HNMR Spectrumof A₂

Mass Spectrum analysis

The mass spectrum of A₁ (Fig 7) reveals the molecularion peak at m/z398 with relative abundance 21% which is agreement with the fromula $C_{19}H_{18}N_4O_4S$ and is equivalent to its molecular weight. The base peak at m/z199.2 this maybe attributed to unusual double chargeion m/2z[21]. The fragment pattern of some importantions are illustrated in scheme 3.



Figure 7: Mass spectrum of A₁







Scheme 3: Fragmentation mechanism of A₁

Molar conductance

The molar condactance of the complexes ZnA_1 and CuA_1 showing that they are non-electrolyte [2]. A value of ($110hm^{-1} cm^2 mol^{-1}$) for ZnA_1 and ($3 ohm^{-1} cm^2 mol^{-1}$) for CuA_1 .



Thermal analysis

Thermal analysis (TG and DTG) of the complexes were used to get information about the thermal stability as well as to verify the status of water molecules into inside or outside the inner coordination spher. The mass losses obtained from TG curves are in a good agreement with the therotical values. Cu complex undergoes decomposition in four steps. The first decomposition step within the range 25-138°C (DTG 110°C) is accompanied by weight loss (obs 1.977%. cal 1.975%) with assigned to the loss one lattice water molecule [2,10]. The second step shows loss in weight within the temperature range of 220-325°C (DTG_{MAX}282°C) which is due to loss C₅H₅N₂ (obs 10.16%.cal10.41%) The third decomposition step begain at 330 and end at 500°C (DTG_{max} 374°C) and represents the decomposition of the sulfa moieties. The residue after the fourth step until 800°C could be identified as C₂₆H₂₀N₂O₂Cu (obs 51% cal.50.72%). TG curve of Zn(II) complex shows a three decomposition steps, the first step at 25-110°C (DTG 77°C) assigned to the loss of two water lattice molecules (obs 4.12 % cal 4.01 %) The second step lies in the temperature rang 250-410°C (DTG_{max} 344) correspods to the loss of C₁₀H₁₂N₆S₂O₄. (obs 34.63% cal 32.43%). The third step begain within 420°C and the (DTG_{max} 521°C) where the ligand decompose in fast rate until 800°C. Fig (8)



Fig 8 : TG and DTG curve of complex CuA₁

Antimicrobial activity

Bacterial sensitivity of(*staphylococeus aureus*) against perpared chemical compounds (S.M. ,A₁ , A₂ , ZnA₁ and CuA₁) were clarified in Table (2 , The maximum inhibition zone was recorded with β – lactam prepared compound (A₂). It was seventeen milimeter (17mm) with concetration (100mg) and eight milimter (8mm) with concentration (1mg) per1 ml DMSO. While others prepared compound (S.M. A₁ , ZnA₁ and CuA₁) were resist with concentration (1mg) and moderate sensitivity with elevated concentration (10mg, 50mg, 100mg) per 1ml DMSO. The results could be explained that β – lactam compound have active to block termination peptido-glycan crosslinking in G+Ve bacteria like *staphylococcus aureus* [22].



hence, It is possible and use it as synthitic product for antimicrobial activity after cytotoxicity study proff or may be used as external use.

Compound	Diameter of inhibition zone (mm) S.aureus Per 1ml DMSO						
	1 mg	10 mg	50 mg	100 mg			
S . M	R	7	9	10			
A1	R	8	11	15			
A2	8	12	15	17			
ZnA1	6	9	12	15			
CuA1	6 9 10 14						

Table 2: Antimicrobial activity of chemical compound toward S. aureus

As for the standard antibiotics, The drugs (Ciprofloxacin, Clarithromycin and Novobiocin) were showed results concrete inhibition zone , while other drugs will be never. Table (3).

Standard Drug	Diameter of inhibition zone (mm)			
	Con.	Abr.	S.aureus	
Cephalexin	30mcg	CL	R	
Ciprofloxacin	5mcg	Сір	20	
Oxacillin	1mcg	Ox	R	
Cloxacillin	1mcg	Cx	R	
Aztreonam	30mcg	ATM	R	
Ampicillin	10mcg	AM	R	
Clarithromycin	15mcg	CLR	23	
Novobiocin	30mcg	NV	24	

Table 3: Antimicrobial activity of standard drug toward *S.aureus*

On the other hand, The ohers three G-Ve clinical bacterial isolates (*E.coli*, *Ps.aerugiosa* and pr.spp.) were highly sinsetive toward shiff-complexes. The maximam inhibition zone were showed by (ZnA_1, CuA_1) against these bacteria in 100mg concentrationper(1ml DMSO), but in the concentration 1mg per (1ml DMSO) the chemical compound had less antimicrobial activity and elevated with 10mg and 50mg concentration. As a result of various metal complexes whith bi- and tridentate Schiff bases containing nitrogen and oxygen donor atoms play important role in biological system and represent interesting models for metalloenzymes ,which efficiently catalyze the reduction of dinitrogen and dioxygen ,In addition to the enhanced activity of the complexes may be ascribed to the increased lipophilic nature of the complexes arising due to chelation^(,), especially Zn(II) complexes. Tabl : 4, Table : 5, Table : 6



Compound	Di	iameter of inl E.coli Pe	hibition zone er 1ml DMSO	(mm)
	1 mg	10 mg	50 mg	100 mg
S . M	R	6	8	11
A1	6	10	12	14
A2	6	10	13	15
A4	7	10	14	17
A6	8	12	14	17

Table 4: Antimicrobial activity of chemical compounds toward E.coli

Table 5: Antimicrobial activity of chemical compoundstoward Ps.aeruginosa

Compound	Diameter of inhibition zone (m	m) Ps.aeru	ginosa Per 1n	nl DMSO
	1 mg	10 mg	50 mg	100 mg
S . M	R	6	7	9
A1	R	7	9	11
A2	6	10	12	12
A4	R	7	12	16
A6	R	8	11	13

Table 6: Antimicrobial activity of chemical compounds toward Pr.ssp.

Compound	D	iameter of inl Pr.ssp	hibition zone . Per 1ml DN	(mm) 1SO
	1 mg	10 mg	50 mg	100 mg
S . M	R	6	7	9
A1	R	8	9	10
A2	R	7	10	12
A4	6	10	12	16
A6	6	9	11	13

The standard antibiotics were showed ineffective toward these bacteria except (Ciprofloxacin , Clarithromycin ,Novobiocin) in *E.coli* and (Ciprofloxacin ,Novobiocin) in *Ps.aeruginosa*as well as (Ciprofloxacin ,Aztreonam ,Novobiocin) in *Pr.spp*.Table : 7 , Table : 8 , Table : 9



Standard Drug	Diameter of inhibition zone (mm)			
Stanuaru Drug	Con.	Abr.	E.coli	
Cephalexin	30mcg	CL	R	
Ciprofloxacin	5mcg	Сір	28	
Oxacillin	1mcg	Ox	R	
Cloxacillin	1mcg	Cx	R	
Aztreonam	30mcg	ATM	R	
Ampicillin	10mcg	AM	R	
Clarithromycin	15mcg	CLR	16	
Novobiocin	30mcg	NV	7	

Table 7: Antimicrobial activity of standard drug toward E.coli

Table 8: Antimicrobial activity of standard drug toward Ps.aeruginosa

Standard Drug	Diameter of inhibition zone (mm)			
Stalluaru Drug	Ps.aeruginosa	Abr.	Con.	
	30mcg	CL	R	
Cephalexin	5mcg	Cip	17	
Ciprofloxacin	1mcg	Ox	R	
Oxacillin	1mcg	Cx	R	
Cloxacillin	30mcg	ATM	R	
Aztreonam	10mcg	AM	R	
Ampicillin	15mcg	CLR	R	
Clarithromycin	30mcg	NV	14	

Table 9: Antimicrobial activity of standard drug toward Pr.ssp.

Ctondord Drug	Diameter of inh	ibition zone (r	nm)
Standard Drug	Con.	Abr.	Pr.
Cephalexin	30mcg	CL	R
Ciprofloxacin	5mcg	Cip	26
Oxacillin	1mcg	Ox	R
Cloxacillin	1mcg	Cx	R
Aztreonam	30mcg	ATM	24
Ampicillin	10mcg	AM	R
Clarithromycin	15mcg	CLR	R
Novobiocin	30mcg	NV	20

REFERENCES

- [1] Saul Pasti. The chemisrty of C=N, first published 1970, John Wiley and Sons Ltd.
- [2] Ebrahimi HP, Hadi JS, Abdulnabi ZA, and Bolandnazar Z. Spectrochim Acta Part A : Mol Biomol Spectr 2014;117 :485-492.
- [3] Gupta KC, Sutar AK, and Lin C. Coord Chem Rev 2009;253: 1926-1946.

RJPBCS



- [4] Hadi JS, and Jarallah HM, Res J Pharm Biol Chem Sci 2013;4(1):292-301.
- [5] Mishra AP, and Gupta P. J Chem Pharm Res 2011;3 (2) : 150-161.
- [6] Alsalim TA, Hadi JS, Ali ON, Abbo HS, and Titinchi SJJ. Chem Central J 2013;7 (3):21-26.
- [7] Alsalim TA, Hadi JS, Al-Nasir EA, Abbo HS, and Titinchi SJJ. Catal Lett 2010;136:228-233.
- [8] S Arulmwugan, HP Karitha and BR Venkatraman. Rasayan J Chem 2010;3(3): 385-410.
- [9] K Singh and D Pal. J Serb Chem Soc 2012;75(7) :917-927.
- [10] ZH Chohan, H Perver, A Rauf and CT Supuran. Metal Based Drugs 2002;8(5): 263-267.
- [11] ST Wadher, MP Puranik, NA Karande and PG Yeole. Int J Pharm Tech Res 2009;1(1)22-33.
- [12] A Kar. Pharmacognosy and Pharmacobiotechnology, 2nd edition , New AG International Limited Puplishers .
- [13] AW Baueyer and WN Kirby. Am J Clin Pathol 1966;45 :493-496.
- [14] JH Zar. Biostatistical Analysis, 5th Edition , 2010 Puplished by Prentice HALL ERRATA / CORRECTIONS .
- [15] Joshi R, Ronad PM, Rayaji A, APandagale A, and Maddi VS. Asian J Pharm Sci Clin Res 2012;2(2):1-9.
- [16] Revanasiddappa BC, Subrahmanyam EVS, and Satyanarayana D. Int J Chem Tech Res 2010;2(1):129-132.
- [17] Bhatk L, Mishra SK, James JP and Shastry CS. J Chem Pharm Res 2010; 3(3):114-118.
- [18] Sharma MC, Sahu NK, Kohli DV, Sharma SCC. Digest J Nnomaterial Biostructures 2009;4 (2):361-367.
- [19] Hesse M, Meier H and Zeeh B. Spectroscopic Methods in Organic Chemistry 6th edition, Thieme . Stuttgart 2002.
- [20] Novkovic SB, Bogdanovic GA, Fraisse B, Ghermani NE and Bouhmaida N. J Phy Chem A 2007;3 (51) :13492.
- [21] Sparkman OD, Penton Z, and Kitson G. Gas Chromotography and mass spectrometry : A practical Guide , 2nd edition, Elsevier 2011.
- [22] Faikong K, Schneper, and Mathee K. J Compilation APMIS 2009.
- [23] BK Singh, D Adhikari, Int J Basic Appl Chem Sci 2012;2 (1) :84-107.