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Isolation and Identification of two bioactive compounds from basidiomycetes fungus Coprinus sp.

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Abstract

The fungus *Coprinus* sp. was isolated and cultivation in laboratory on two PDA medium with some growth induction .Two bioactive chemical compounds were isolated and purified from mycelial culture of *Coprinus* sp. by using potato dextrose agar and potato dextrose broth . The two compounds were identified by using GC-mass technique .The molecular weight of purified compounds 1 and 2 were 352 KD and 388 KD respectively and chemical formula of compound 1 is C21H24N2O3 which isolated from solid medium while compound 2 is C24H26O4 which isolated from broth medium . The antibacterial activity of the purified compounds against three bacterial species *E. coli* , *Proteus* sp. and *S. aureus* were tested by using a disk diffusion agar method reaching to 40 , 30 , 20 for compound 1 and 37, 25 , 17 mm for compound 2.

Key wards: Isolation, Identification, Bioactive comp., Coprinus sp.

عزل وتشخيص اثنين من المركبات الفعالة من الفطر البازيدي. Coprinus sp

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الخلاصة

عزل الفطر البازيدي Coprinus sp وزرع مختبريا على وسط PDA ياستخدام محفزات نمو . تم عزل وتتقية اثنين من المركبات الكيميائية الفعالة من مزرعة الفطر باستخدام وسط البطاطا والدكستروز الصلب ووسط البطاطا و الدكستروز السائل . شخص المركبان باستخدام تقنية طيف الكتلة C21H24N2O3 (1) . شخص المركبان باستخدام تقنية طيف الكتلة GC-mass . الوزن الجزيئي للمركبين1 و 2 بلغ 38 kd على التوالي . الصيغة الكيميائية للمركب الأني C24H26O4 (2) والمعزول من الوسط السائل . أختبرت الفعالية الحيوية للمركبين وقد بعض الاتواع البكتيرية E. coli.، S. aureus و 20 مليمتر للمركب الأول و 37 ضد بعض الاتواع البكتيرية للمركب الثاني (2) على التوالي .

كلمات المفتاح: عزل، تشخيص ، مركبات فعالة ، الفطر البازيدي.

Introduction

The researches interest to explore new antimicrobial agents from fungi is continued ,fungi are a rich source of bioactive secondary metabolites and mushroom –forming fungi are especially known for the synthesis of numerous bioactive compounds (Anke et al., 2004; Keller et al., 2005; Agger , 2009; Muhsin et al., 2011).

Coprinus belongs to mushrooms , which is a black spored family coprinaceae (all species go through an auto digestion at maturity in which the cap forms black spores). The genus Coprinus is considered to be more than 100 species distributed from the northern hemisphere to south African (Keirle *et al.*, 2004). The classification of the *Coprinus* species is still unclear because it was only based on morphological

characteristics without molecular analysis (Mwita et al., 2010). Extracts from several members of the genus Coprinus are known in many part of the world to exhibit among others, antimicrobial, antitumor, hypoglycemic, antinematodes and antioxidant effects. (Ndyetabura et al., 2010). Examples of the bioactive compounds that have been reported present in Coprinus mushrooms extracts are : abroad spectrum bioactive indole compound tryptamine (Worthen et al., 1962), Miaceol (Asterol) with antibacterial activity against bacteria Corvnebacterium xerosia the Staphylococcus aureus (Zahid et al., 2006) and (2,2)-4-oxo-2,5- heptadienedioic acid, which has inhibitory activity against glutathione S- transferase an enzyme that have been implicated in the resistance of cancer cells against chemotherapeutic agents especially alkylating drugs (Zahid et al., 2006). According to our knowledge so far a little information is available about the production of secondary bioactive metabolites by this fungus, this report elucidates interesting chemical compound, extracted, purified and identified from mycelial culture of Coprinus sp. As a bioactive agents tested against a selected species of bacteria.

Material and Methods

1: Fungal Mushroom culture

Fruiting bodies were collected from palm trees (Phoenix dactylifera) southern of Iraq . In the laboratory , small Pieces ($0.5~\rm cm$ long) were cut from the fruiting body surface , sterilized with 10% sodium hypochlorate for 3 min. , washed with sterile distilled water and placed on potato dextrose agar (PDA) with growth indicated (Aspargine and thiamine) in petri dishes , plates were incubated at $25\pm1~\rm Co$ for two weeks . After cultivation , the mycelium was removed from the agar medium surface and amended into a liquid culture medium potato dextrose broth (PDB) in 1 L conical flasks and incubated at 25 Co in a rotary shaker incubator for 3 weeks .

2: Extraction, Isolation and Purification

The fungal culture in broth medium was filtered on Watmann No. 1 filter paper , the filtrate was extracted three time with ethyl acetate ($1{:}1~\text{v/v}$) using separating funnel , while fungal culture on the solid media prior to extraction with ethylacetate,the solid medium were diluted with H2O and blended using blender (Zur , 2001).Organic layer was collected and dehydrated with Na2SO4 then placed in Petri dishes and dried at room temperature , thin layer chromatography (TLC) was applied for the isolation of extracted metabolites using silica gel of $2{\times}10~\text{cm}$ (Silica gel

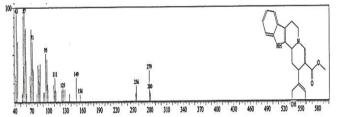
GF2 Merck) and Rf value (Rate flow) were measured . Purification of extracted compounds was made on silica gel column chromatography (silica gel mesh 60) (Column 1.5×50 cm and elution with methanol - ethyl acetate (1:1). a further purification of fraction compound were made by using another column 1.5×50 cm and using eluent cyclohexane and ethyl acetate (1: 1) . The identification of the purified compounds was made by using Gas chromatography GC-Mass technique type (Simadzu . GCMS-QP2010 Ultra)

3: Bioactive test

Disc diffusion agar method (Casals, 1979) was used to examine the antimicrobial activity of the purified compounds, Three strains of pathogenic bacteria from (Lab. of Pharmacy college), Escherichia coli , Proteus sp. and Staphylococcus aureus were used for this purpose . 2.5 mg of the dried fungal extract was dissolved in 1 ml of dimethyl sulfuoxide (DMSO) solvent used as stock solution test . Disk of 0.6 mm diameter Whatmann for this No. 1 filter paper was sterilized and soaked in the fungal extract solution and placed on plates containing Muller -Hinton agar (MHA) medium inoculated with 0.1 ml suspension of bacterial strains by streaking method. The test was carried out in triplicates . Antimicrobial activity was assayed by measuring the inhibition zone around the disk in mm.

Results:

The fungus Coprinus sp. Which isolated in this study has some characters such as: Basidiocarp brown in color, brown cap with brown -black spores (4.5 - $5.5 \times 4 - 7.5$) µm stalk diameters (5×0.25) cm and spores are brown in color. Mycelium white in color when grow on the solid media and grow up on the Two compounds were isolated and media surface purified from solid and broth culture of Coprinus sp. Mycelium. The compound (1) isolated from solid culture, while compound (2) isolated from broth culture . Based on Gas chromatography (GC-mass) apparently that the molecular formula of compound (1) is C₂₁H₂₄N₂O₃with molecular weight 352Kd and its chemical name is 2-(1-hydroxymethyl - propenyl) -1,2,6,7,12,12b-hexahydro-indo [2,3-a] quinolizine -3carboxylic methyl ester showed in fig (1), While the molecular formula of compound (2) is $C_{24}H_{36}O_4$ and its name is Cycloheptylnonphthallate with molecular weight 388Kd fig (2). The purified compound 1 and 2 appeared antibacterial activity against bacterial species. The inhibition zone diameter of compound (1) reach to (40, 30, 20) mm for *S. aureus*, *E. coli* and *Proteus* sp. respectively and (37, 25, 17) mm of compound (2) for three bacterial species respectively fig (3).



Fig(1): Chemical structure and GC –Mass spectroscopy of compound **-1** (2-(1-hydroxymethyl-propenyl)-1,2,6,7,12,12b-hexahydro-indo [2,3-a] quinolizine -3-carboxylic methyl esteIsolation from solid media

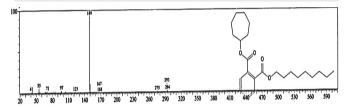


Fig (2): Chemical structure and GC –Mass spectroscopy of compound- **2**(Cycloheptylnonphthalate) isolation from broth media

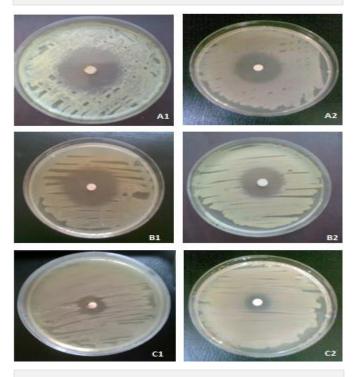


Fig (3): Inhibition zone diameter (mm) :- 1: compound
No. (1) and 2: Compound No(2)

A = E. coli B= S. aureus C= Proteus sp.

Discussion

Fungi in general and mushroom in particular are a good source for antimicrobial products (Janes et al., 2007) . Results showed the activity of two compounds purified from *Coprinus* sp. (1,2) against gram positive and gram negative bacteria, so this activity may be due to active substances present in the *Coprinus* sp. tissues (Ndyetabura et al., 2010), also these results agreed with Zenkora et al., (2003) whom reported that the Coprinus representatives produces compounds which are able to inhibit growth of all common gram-negative and gram positive bacteria and fungal strains .The antimicrobial activity of compound (1)against bacterial strains may be due to the quinolizine and indole group in its structure, also this groups have different biological activity such as antimicrobial, antiinflamatory, antituberculosis, cytotoxicity (Thakur et al., 2010), while the antimicrobial activity compound (2) may be due to its one of the phthalide derivatives which are compounds of polyketide metabolism, they are produced by a wide range of organisms, i.e marine and terrestrial fungi (Almedia et al., 2011) . Phthalides exhibit equaly broad spectrum of bioactivity, including modulation of central nervous system protection against brain eschemia, as well as antibacterial, antifungal and phtytoxic activity. Moreover, the rings compounds acts as a protoplasm toxin to destroy the cell wall system and to precipitate protein in cells (Gayon, 1972). The compound produced from solid media have more biological activity against bactetria than liquid media, several researches showed that produced through solid media are more stable and produced in higher quantities than liquid. In this fermentation technique, the substrate can be used for long fermentation periods, hence, this technique supports controlled release of nutrients (Subramaniyam and Vimala, 2013). The role of the physiological and genetic properties of the microorganism used during growth on solid substrates compared with aqueous solution has so for been all but neglected ,despite the fact that it may be the microbiology that makes solid state fermentation advantageous against submergied fermentation biotechnology (Acura-Arguelus et al., 2004). The fact basidiomycetes have been insufficiently investigated coupled with broad range of structural types of antibiotics. However basidiomycetes may become source of new and useful bioactive compound (Srivastava and Sharma, 2011). To our knowledge, no investigation has been performed for comparing antimicrobial activity potential of basidiomycetes strains. Further studies on isolation and identification of the active compounds may provide a better source for developing new therapeutic agents.

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